

## HiYield™ 96-Well G-50 Cleanup Kit

Cat. No.:	YCG96B-2	YCG96B-4	YCG96B-10
<b>Product Name:</b>	HiYield™ 96-Well G50 Cleanup Kit		
<b>Reactions:</b>	2	4	10
<b>DNA Size Range:</b>	Larger than 20 bases		
<b>Format:</b>	96-Well Plates		
<b>Operation:</b>	Centrifuge		
<b>Operation Time:</b>	10 Minutes		

### Introduction

HiYield™ 96-Well G50 Cleanup Kit is ideal for removing excess dye terminator, freeing nucleotides from sequencing and labeling reactions, desalting and buffer exchange. G50 plates can purify DNA fragments larger than 20 bases in length with low molecular weight material retained in the gel matrix. Since G50 plates are designed to purify DNA fragments larger than 20 bases only, they are not recommended for PCR product primer removal.

### Features

Entire procedure could be completed within 10 minutes.

Ready-to-use prehydrated gel-filtration 96-well plates.

Optimized for efficient removal of any dye terminator.

### Contents

ITEM	YCG96B-2	YCG96B-4	YCG96B-10
G50 Plate	2 plates	4 plates	10 plates
350ul Collection Plate	2 plates	4 plates	10 plates

### Applications

Desalting, Dye Terminator Removal, Buffer Exchange.

### Quality Control

The quality of HiYield™ 96-Well G50 Cleanup Kit is tested on a lot to lot basis. The particle size and quality is tested. The purified DNA is checked by electrophoresis.

### Things Before Starting:

In the event of gel drying (cracking), add 50-100  $\mu$ l of ddH<sub>2</sub>O to each well of the HiYield™ 96-Well G50 Cleanup Kit before use. The optimal sample volume is 20 to 50  $\mu$ l (50  $\mu$ l maximum).

### Purification/Desalting Protocol

<b>Step 1</b>	<ul style="list-style-type: none"> <li>★ Remove the adhesive film from the <b>G-50 Plate</b>.</li> <li>★ Place the <b>G-50 Plate</b> on a 2 ml collection plate and centrifuge at 2,500 x g for 5 minutes.</li> </ul>
<b>Step 2</b>	<ul style="list-style-type: none"> <li>★ Transfer the <b>G-50 Plate</b> to a <b>350 <math>\mu</math>l Collection Plate</b>.</li> <li>★ Carefully load the sample (20-50 <math>\mu</math>l) onto the center of each gel bed surface.</li> </ul>
<b>Step 3</b>	<ul style="list-style-type: none"> <li>★ Centrifuge again at 2,500 x g for 5 minutes.</li> <li>★ Each purified sample can be recovered at the bottom of the <b>350 <math>\mu</math>l Collection Plate</b> (approximately the same volume as the loaded sample).</li> </ul>

### Buffer Exchange Protocol

<b>Step 1</b>	<ul style="list-style-type: none"> <li>★ Remove the adhesive film from the <b>G-50 Plate</b>.</li> <li>★ Place the <b>G-50 Plate</b> on a 2 ml collection plate and centrifuge at 2,500 x g for 5 minutes.</li> </ul>
<b>Step 2</b>	<ul style="list-style-type: none"> <li>★ Discard the flow-through in the 2 ml collection plate and place the <b>G-50 Plate</b> back on the same 2 ml collection plate.</li> <li>★ Add 350 <math>\mu</math>l of desired buffer to each well of the <b>G-50 Plate</b> and centrifuge at 2,500 x g for 5 minutes.</li> </ul>
<b>Step 3</b>	<ul style="list-style-type: none"> <li>★ Transfer the <b>G-50 Plate</b> to a <b>350 <math>\mu</math>l Collection Plate</b>.</li> <li>★ Carefully load the sample (20-50 <math>\mu</math>l) onto the center of each gel bed surface.</li> <li>★ Centrifuge again at 2,500 x g for 5 minutes. Each purified sample can be recovered at the bottom of the 0.35 ml collection plate (approximately the same volume as the loaded sample).</li> </ul>